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(54) HUMAN TAKI DNA ENCODING THE SAME

(57) A TGF- β -activated kinase comprising an amino acid sequence from Met at position 1 to Ser at position 579 in the amino acid sequence as set forth set forth in SEQ ID NO: 5, and DNA encoding the kinase.

EP 0 919 621 A1

Description

Technical Field

5 [0001] The present invention relates to a kinase (transforming growth factor- β (TGF- β) activated kinase 1; TAK1) carrying the signal transduction system of the TGF- β family, said kinase being activated by TGF- β , a method for producing the kinase, and a human gene encoding the kinase. TAK1, also referred to as the activator of MAPK kinase (AMK-1), is an enzyme that is activated by TGF- β and BMP (bone morphogenetic protein) and that phosphorylates and thereby activates the MAPK kinase.

10 Background Art

[0002] Receptors of the TGF- β superfamily contain Ser/Thr kinase in the intracellular domain and are divided into type I that has a repeated sequence of Gly and Ser (GS box) at the amino-terminal end near the transmembrane domain and type II that does not have the GS box. It is believed, in the case of TGF- β , that it forms a complex with the type I receptor after a ligand has bound to the type II receptor, in which a constitutively phosphorylated kinase of the type II receptor phosphorylates the vicinity of GS box of the type I receptor, thereby activating the type I receptor so that a signal from the above ligand is transmitted into the cell. However, little is known about signaling molecules downstream of this receptor.

20 [0003] For a eukaryotic budding yeast *Saccharomyces cereviceae*, it is known, as a signal transduction cascade in which an extracellular mating pheromone causes conjugation, that the mating pheromone activates a G-protein, which in turn activates a MAPKK kinase (MAPKKK) (Ste11), and the activated MAPKKK phosphorylates and activates a MAPK kinase (MAPKK), and then the thus activated MAPKK (Ste7) phosphorylates and activates a MAP kinase (mitogen-activated protein kinase; MAPK), and finally the MAPK activates FUS1 protein to initiate cell conjugation.

25 [0004] As such a MAPKKK, TAK1 (TGF- β activated kinase 1) derived from mouse has been known so far (K. Yamaguchi et al., Science (1995) 270, 2008-2011).

Disclosure of Invention

30 [0005] The present invention intends to provide a novel factor in the signaling system of the receptor of mammalian TGF- β said factor being located downstream of said receptor and being involved in said signaling system; gene encoding said factor; and a method for producing said factor.

[0006] In efforts to solve the above problems, the applicants of the present invention have succeeded in inserting a human-derived cDNA into a yeast *Saccharomyces cereviceae* that is deficient in MAPKKK activity (Ssk2/Ssk22, Sho1) 35 in the signal transduction cascade of the above mating pheromone, screening for cDNAs that can complement the activity-deficient MAPKKK, and cloning cDNAs that can complement the activity-deficient MAPKKK, and thereby have accomplished the present invention.

[0007] Thus, the present invention provides a polypeptide having a kinase activity that is activated by transforming growth factor (TGF)- β , said polypeptide comprising an amino acid sequence from Ser at position 23 to Ser at position 40 579 set forth in SEQ ID NO: 5.

[0008] The present invention also provides a polypeptide having a kinase activity that is activated by TGF- β , said polypeptide comprising an amino acid sequence from Met at position 1 to Ser at position 579 set forth in SEQ ID NO: 5.

[0009] The present invention also provides DNA encoding a polypeptide having a kinase activity that is activated by TGF- β , said polypeptide comprising an amino acid sequence from Ser at position 23 to Ser at position 579 set forth in 45 SEQ ID NO: 5.

[0010] The present invention also provides DNA encoding a polypeptide having a kinase activity that is activated by TGF- β , said DNA comprising a nucleic acid sequence from T at position 249 to A at position 1919 set forth in SEQ ID NO: 5.

[0011] The present invention also provides DNA encoding a polypeptide having a kinase activity that is activated by 50 TGF- β , said polypeptide comprising an amino acid sequence from Met at position 1 to Ser at position 579 set forth in SEQ ID NO: 5.

[0012] The present invention also provides DNA encoding a polypeptide having a kinase activity that is activated by TGF- β , said DNA comprising a nucleic acid sequence from A at position 183 to A at position 1919 set forth in SEQ ID NO: 5.

55 [0013] The present invention also provides a vector comprising any of the above-mentioned DNAs, a host cell transformed with a vector comprising any of the above-mentioned DNAs, and a method for producing a polypeptide having a kinase activity that is activated by TGF- β , which method comprises culturing a host cell transformed with a vector comprising any of the above-mentioned DNAs and then recovering a product from the culture.

[0014] The present invention also provides a polypeptide having a kinase activity that is activated by TGF- β , said polypeptide being produced by the above method, and a kinase that is activated by TGF- β , said kinase comprising an amino acid sequence from Ser at position 23 to Ser at position 579 set forth in SEQ ID NO: 5.

[0015] The present invention also provides a fusion protein of the above polypeptide, protein and another protein.

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Brief Explanation of the Drawings

[0016]

- 10 Fig. 1 shows a yeast expression vector pNV11.
Fig. 2 is a graph showing the effects of TGF- β addition on the expression of various TAK1 genes evaluated using a luciferase gene as a reporter gene.
Fig. 3 is a graph showing the effects of TGF- β and BMP-4 on the activity of a TAK1 gene in MC3T3-E1 cells as measured by an immunoprecipitation method and a coupled kinase method.
15 Fig. 4 is a graph showing the effects of various concentrations of TGF- β or BMP-4 on TAK1 kinase activity in the cells transfected with the HA-TAK1 gene. TAK1 Δ N shows a result when the cells transfected with the TAK1 Δ N gene were not stimulated by either TGF- β or BMP-4.
Fig. 5 shows a comparison of the base sequence of DNA encoding mouse TAK1 and that of DNA encoding human TAK1.
20 Fig. 6 shows a comparison of the base sequence of DNA encoding mouse TAK1 and that of DNA encoding human TAK1.
Fig. 7 shows a comparison of the base sequence of DNA encoding mouse TAK1 and that of DNA encoding human TAK1.
Fig. 8 shows a comparison of the base sequence of DNA encoding mouse TAK1 and that of DNA encoding human TAK1.
25 Fig. 9 shows a comparison of the base sequence of DNA encoding mouse TAK1 and that of DNA encoding human TAK1.
Fig. 10 shows a comparison of the amino acid sequence of mouse TAK1 and that of human TAK1.
Fig. 11 shows a comparison of the amino acid sequence of mouse TAK1 and that of human TAK1.

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Embodiment for Carrying out the Invention

[0017] In accordance with the present invention, the cloning of the desired gene can be detected by inserting, for example, an expression vector comprising a mammalian cDNA into a yeast that is deficient in the MAPKKK activity and has a readily detectable reporter gene at the terminal of the cascade, and then by expressing said reporter gene to detect the introduction of cDNA that complements the deficient MAPKKK activity. Moreover, another yeast can be used that is deficient in Ssk2/Ssk22 and Sho1 activity and that functions, for example, in a high osmotic pressure-signaling system.

[0018] As such a detection system, there can be used a MAPK route that transmits a signal of the mating pheromone in *Saccharomyces cerevisiae* (I. Herskowitz, Cell, Vol. 80, 187 (1995); D.E. Lein et al., Curr. Opin. Cell Biol. Vol. 7, 197 (1995); J. Schulz et al., Curr. Opin. Gene Dev., Vol. 5, 31 (1995)). The normal signal transduction cascade in this system consists of Ste11 kinase, Ste7 kinase, and Fus3/Kss1 kinase, which correspond to MAPKKK, MAPKK, and MAPK, respectively. Ste11, Ste7, and Fus3/Kss1 sequentially act to thereby transmit signals to the transcription factor Ste12, which Ste12 in turn activates the transcription of a mating-specific gene such as FUS1.

[0019] With respect to the screening of cDNA, there can be used a cascade that has a functional mutation of Ste7 (STE7^{P368}) and a deficiency mutation of Ste11 (Ste11 Δ) in the above cascade (K. Irie et al., Science Vol. 265, 1716 (1994)). In this system, it has been confirmed that a mammalian Raf or an activated type of MEKK (Fat Δ N or MEKK Δ N, respectively) complements the deficiency of Ste11 activity in a Ste7^{P368}-dependent manner, when it is monitored by a histidine phenotype (His) imparted by the reporter gene FUS1p::HIS3 corresponding to the mating route. Thus, by introducing the subject cDNA into a yeast having the above mutated cascade and by detecting the histidine phenotype, cDNA that can complement Ste11 Δ (MAKK deficiency) can be selected.

[0020] As the subject cDNA, there can be used a cDNA library of any mammalian origin. For example, a cDNA expression library from a mouse cell line such as the mouse cell line BAF-B03. This cDNA library can be obtained by cloning cDNA corresponding to poly (A)-RNA from a mouse IL-3-dependent pro- β cell line BAF-B03 under the control of TDH3 promoter of a yeast expression vector pNV11. Another example of the subject cDNA library to be used is a human cell line such as a cDNA expression library from a human cell line Jurkat.

[0021] One positive clone was obtained by screening the above cDNA library using the above screening system. The base sequence of cDNA of this clone and the amino acid sequence encoded by the cDNA correspond to the nucleotide

EP 0 919 621 A1

numbers 223 to 1893 and the amino acid numbers 23 to 579 of SEQ ID NO: 1.

[0022] cDNA libraries from a human cell line can be screened by the above screening system. Alternatively, cDNA libraries from a human cell line can be screened using a mouse cDNA as a probe in the manner described above.

5 [0023] The cDNA of another positive clone and the amino acid sequence encoded by the cDNA correspond to the nucleotides 249 to 1919 and the amino acids 23 to 579 set forth in SEQ ID NO: 5.

[0024] In order to obtain a longer cDNA (full-length cDNA), the above cDNA libraries were screened using said cDNA as a probe to thereby obtain multiple positive clones. These clones had a 5'-extension of approximately 230 bp to said cDNA. cDNA that contains this 5'-extension was designated as TAK1 cDNA and cDNA that was cloned first and that does not contain this 5'-extension was designated as TAK1ΔN cDNA. The nucleotide sequence of TAK1 cDNA is shown 10 in 1 to 2443 of SEQ ID NO: 1, and the amino acid sequence encoded by the cDNA is shown in the amino acid numbers 1 to 579 of SEQ ID NO: 1. The protein or the polypeptide represented by the amino acid sequence is designated as TAK1 protein or polypeptide. In contrast, the protein or the polypeptide represented by the amino acid sequence encoded by TAK1 ΔN cDNA is designated as TAK1 ΔN protein or polypeptide. Furthermore, the nucleotide sequence 15 of human TAK1 cDNA is represented by the nucleotides 1 to 2656 and the amino acid sequence encoded by it is represented by the amino acids 1 to 579 of SEQ ID NO: 5.

[0025] The primary sequence of the TAK1 protein suggests that this protein has a protein kinase catalysis domain at the N-terminal end and a C-terminal domain of about 300 amino acid residues. This catalysis domain contains a consensus sequence corresponding to the protein kinase subdomains I to XI (S.K. Hanks et al., Science 241, 42 (1988)). This catalysis domain has an about 30% identity with the amino acid in the Raf-1 (T.I. Bonner et al., Nucleic Acids Res. 20 Vol. 14, 1009 (1986)) and MEKK (C.A. Langer-Carter et al., Science Vol. 260, 315 (1993)). The sequence of the 300 amino acid residues at the C-terminal following the above catalysis domain does not have a conspicuous homology with other proteins.

[0026] TAK1 ΔN cDNA deficient in the codons of 22 amino acids at the N-terminal which has been introduced into a yeast having a ste11Δ mutation can complement the ste11Δ mutation (MAPKKK deficiency), but the full-length TAK1 cDNA introduced into a ste11 Δ mutant does not complement the ste11Δ mutation. It is believed, therefore, that TAK1 kinase is activated by the removal of 22 the amino acids in the N-terminal.

[0027] Thus, the present invention provides DNA encoding a polypeptide comprising an amino acid sequence from Met at position 1 to Ser at position 579 set forth in SEQ ID NO: 5. This DNA includes, as a typical example, DNA encoding a polypeptide comprising an amino acid sequence from the amino acid Ser at position 23 to the amino acid Ser at position 579, and DNA encoding a polypeptide comprising an amino acid sequence from an amino acid Glu at position 30 to an amino acid Asp at position 295. However, the DNA of the present invention is not limited to the above DNAs and may also include DNA encoding a polypeptide comprising an amino acid sequence from any of the amino acids from Met at position 1 to Glu at position 30 to the amino acid Asp at position 295.

[0028] Because it is believed that even from DNA encoding a polypeptide having an extended N-terminal, an active enzyme could be obtained by processing of the polypeptide after expression, and that it will have a similar kinase activity even without regions other than the kinase at the C-terminal.

[0029] The present invention also provides polypeptides or proteins, especially polypeptides or proteins retaining the TAK1 activity, having the amino acid sequences corresponding to the nucleotide sequences of various DNAs mentioned above. As a more specific example, the present invention relates to polypeptides or proteins expressed by introducing 40 various DNAs mentioned above as they are inserted into, for example a vector, especially an expression vector, into host cells such as animal cells or microorganism cells.

[0030] Typically, the polypeptide or the protein of the present invention has an amino acid sequence from any of an amino acid from Met at position 1 (inclusive) to Ser at position 23 (inclusive) to the amino acid Ser at position 579 of SEQ ID NO: 5.

[0031] The present invention also provides a fusion protein of the above polypeptide or protein and another protein. Another protein that is fused with a polypeptide or a protein having the TAK1 activity may be chosen as appropriate in addition to hemagglutinin mentioned in examples. DNA encoding a fusion protein of a polypeptide or a protein having the TAK1 activity with another protein can be constructed and expressed by the method set forth in Example 4.

[0032] As mentioned earlier, cDNA encoding human TAK1 can be obtained using cDNA encoding mouse TAK1; Examples 5 and 6 describe the isolation of cDNA encoding human TAK1.

[0033] The various DNAs of the present invention mentioned above can be cloned from an animal cell as, for example, cDNA by the method set forth in Example 2. Mutated or modified DNA as compared to the original cDNA can be prepared using the original cDNA as a template in a conventional method such as PCR amplification and site-specific mutagenesis.

[0034] The polypeptides or proteins of the present invention can be obtained by expressing the corresponding DNA in a suitable host cell. As the host cell in this case, there may be used cultured cells of higher eukaryotic cells such as human, monkey, mouse, hamster, and frog cells, for example THP-1 cells, MC3T3-E1 cells, XTC cells, MvILu cells, CHO cells, and COS cells; cultured cells of lower eukaryotic cells including, for example, fungi such as the fungi of

genus *Aspergillus* such as *Aspergillus niger*; or yeasts including, for example, the yeasts of genus *Saccharomyces* such as *Saccharomyces cereviceae*, and the like. As the host cell, furthermore, prokaryotic cells including, for example, bacteria such as *Escherichia coli* may be used.

[0035] When the desired DNA is to be expressed in these hosts, an expression-regulating sequence such as a suitable promoter is used depending on the host. In the expression in animal cells, for example, a plasmid containing each of promoters such as pCDM8, pSV, and pEP is used, and in yeast hosts, a plasmid such as pNV11 is used, and in *Escherichia coli*, a plasmid such as pGEMEX and pJEX is used.

[0036] Transformed hosts may be cultured in a conventional method. The polypeptides or proteins of the present invention can be produced using a transgenic animal (Glaser, V., SPECTRUM Biotechnology Applications, 1993) and an insect such as silkworm (Maeda, S. et al., Nature (1985) 315, 592-594) as a host. Recovery and/or purification of polypeptides and proteins thus produced may be accomplished by a commonly used method for enzyme purification such as centrifugation, filtration, gel filtration chromatography, and affinity chromatography.

[0037] Kinases activated by TGF- β that carries the signal transduction system of the TGF- β family of the present invention are useful for use in the search of drugs that inhibit or promote signaling of TGF- β and its superfamily known to be involved in various diseases.

Examples

[0038] The present invention will now be explained in further details with reference to the following examples.

Example 1. Construction of cDNA library

[0039] cDNA was synthesized from poly(A)-RNA from a mouse IL-3-dependent cell line BAf-BO3 according to a conventional method and was then inserted into a yeast expression vector pNV11 (Ninomiya-Tsuji, J. et al., Proc. Natl. Acad. Sci. U.S.A. 88, 9006-9010 (1991)) shown in Fig. 1 under the control of the TDH3 promoter to construct a cDNA library.

Example 2. Screening of cDNA library

[0040] A cDNA library prepared in Example 1 was screened using *Saccharomyces cereviceae* SY1984-P (his3 Δ , ste11 Δ , FUS1p::HIS3, STE7P368). In this yeast, Ste11 has been mutated and the activity is deficient in the signal transduction system of mating pheromone, and the FUS1 upstream activation sequence has been ligated to the HIS3 open reading frame to form a reporter gene. The yeast strain is deficient in the original his3 and thus it can grow only when exogenous histidine is present in the culture medium or when the mutation-derived deficiency in the Ste11 activity has been complemented.

[0041] *S. cerevisiae* SY1984-P was transformed with various plasmids. The plasmids used were YCplac22 (vector), pRS314PGKMEKKCT (expresses MEKK Δ N (K.J. Blumer et al., Proc. Natl. Acad. Sci. U.S.A. Vol. 91, 4925 (1994) that lacks the N-terminal domain downstream of the PGK1 promoter), and pADU-Raf Δ N (expresses Raf Δ N (K.Irie et al., Science Vol. 265 1716 (1994) that lacks the N-terminal domain downstream of the ADH1 promoter). These transformants were plated onto a SC-His plate that lacks histidine and were incubated at 30°C. As a result, the yeast that was transformed with the YCplac22 vector did not grow but the yeast transformed with pRS314PGKMEKKCT or pADU-Raf Δ N did. This confirmed that the screening system is effective.

[0042] Then the above screening system yeast strain YS1984-P was transformed with the cDNA library constructed in Example 1 and then was screened on an SC-His plate to obtain one positive clone pNV11-HU11. The cDNA of this clone was designated as TAK1 Δ N cDNA. The nucleotide sequence of this cDNA was determined by the dideoxy nucleotide chain termination method. The nucleotide sequence corresponds to the sequence of nucleotides 223 to 1893 set forth in SEQ ID NO: 1 and the amino acid sequence encoded by it corresponds to Ser at position 23 to Ser at position 579 of the amino acid sequence set forth in SEQ ID NO: 1.

[0043] Then, in order to clone the full-length cDNA, the above TAK1 Δ N cDNA was radiolabeled and used as a probe to further screen the cDNA library obtained in Example 1. Thus, multiple positive clones were obtained. The cDNA of this clone was subcloned into the EcoRI site of pBS vector (manufactured by Stratagene) to obtain pGS-TAK1-5'. This clone was a full-length clone containing the initiation codon ATG. The cDNA was designated as TAK1 cDNA. The nucleotide sequence thereof is shown set forth in SEQ ID NO: 1. The full-length amino acid sequence of Met at position 1 to Ser at position 579 has been encoded in nucleotides 1 to 2443 in the above sequence.

Example 3. Tissue distribution of TAK1 gene

[0044] Total RNA was extracted from various tissues of mice and was subjected to Northern blotting using the above

radiolabeled TAK1 cDNA as a probe to find that RNA that hybridizes with TAK1 cDNA was expressed in all the tissues or organs tested (spleen, thymus, lung, heart, liver and brain). It was present at high levels in the spleen, thymus, and brain, and at low levels in the lung, heart, and the liver.

5 Example 4. Properties of TAK1 kinase

- [0045] In order to study the functions of the kinase activated by TGF- β in mammalian cells, TAK1 cDNA and TAK1 Δ N cDNA were inserted into a mammalian expression vector pEF (H. Shibuya et al., Nature Vol. 357, 700 (1992)) under the control of the human elongation factor (EF) promoter to obtain expression plasmids pEF-TAK1 and pEF-TAK1 Δ N.
- 10 The expression plasmids pEF-TAK1 and pEF-TAK1 Δ N contain a full-length TAK1 coding sequence and TAK1 Δ N coding sequence, respectively, under the control of the EF promoter.
- [0046] Thus, 2.3 kb of Xhol fragment of pNV11-HU11 was inserted into a Xhol gap of PBS to obtain pBS-TAK1 Δ N. pEF-MSS1 (H. Shibuya et al., Nature Vol. 357, 700 (1992)) was cleaved with EcoRI and XbaI, into which were introduced a synthetic EcoRI-Xhol linker (sense strand: 5'-AATTGCCACCATGGC-3') (SEQ ID NO: 2); antisense strand: 15 5'-TCGAGCCATGGTGGCG-3' (SEQ ID NO: 3) (containing the initiation codon ATG), and an Xhol-HindIII fragment and a HindIII-XbaI fragment from pBS-TAK1 Δ N to construct pEF-TAK1 Δ N. PBS was cleaved with EcoRI and Xhol, into which were inserted an EcoRI-SacI fragment from pBS-TAK1 Δ -5' and a SacI-Xhol fragment from pBS-TAK1 Δ N to obtain pBS-TAK1 containing the full-length cDNA of TAK1 (TAK1 cDNA). pEF-MSS1 was cleaved with EcoRI and Sall, into which was inserted an EcoRI-SacI fragment from pBS-TAK1 to construct pEF-TAK1.
- 20 [0047] E. coli having the plasmid pEF-TAK1 has been internationally deposited under the provisions of the Budapest Treaty as *Escherichia coli* MC1061/P3 (pEF-TAK1) and E. coli having the plasmid pEF-TAK1 Δ N as *Escherichia coli* MC1061/P3 (pEF-TAK1 Δ N) on September 29, 1995 with the National Institute of Bioscience and Human Technology, the Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as FERM BP-5246 and FERM BP-5245, respectively.
- 25 [0048] The TAK1 gene contained in the plasmid pEF-TAK1 can be excised using an appropriate restriction enzyme such as EcoRI and BamHI.
- [0049] A study of the effects of TAK1 on the induction of gene expression by various ligands has revealed that TAK1 has an effect on gene induction by TGF- β . An initial cellular response to TGF- β induces an elevation in the mRNA level of plasminogen activator inhibitor 1 (PAI-1) (M.R. Keeton et al., J. Biol. Chem. Vol. 266, 23048 (1991)).
- 30 [0050] In order to study the effects of TAK1 on TGF- β response, TGF- β reporter plasmid p800neoLUC containing a luciferase gene regulated by the PAI-1 promoter induced by TGF- β (M. Abe et al., Analyt. Biochem., Vol. 216, 276 (1994)) was transiently transfected to MvLu lung epithelial cells by the calcium phosphate method (H. Shibuya et al., Nature Vol. 357, 700 (1992)). In this method of measurement, the luciferase activity induced by TGF- β can be measured by the transfection of p800neoLUC onto the MvLu lung epithelial cells. The MvLu cells transiently transfected by 35 p800neoLUC responded to TGF- β with a 4- to 5-fold enhanced reporter gene activity. This result is shown in the vector section of Fig. 2.
- [0051] The previously constructed TAK1 or TAK1 Δ N expression plasmid was transiently transfected into MvLu cells. Expression of TAK1 slightly enhanced the expression of TGF- β -derived genes and TAK1 Δ N constitutively activated the expression of the PAI-1 gene (the section of TAK1 Δ N of Fig. 2). The level of constitutive expression of a reporter gene 40 by TAK1 Δ N is equal to that in a transfectant treated with TGF- β . Thus, the activated TAK1 (i.e. TAK1 Δ N) can transfer signals in the absence of TGF- β . Furthermore, when TGF- β was added to a TAK1 Δ N transfectant, the expression of the PAI-1 gene was further enhanced.
- [0052] In Fig. 2, the open bars represent a case in which no induction of TGF- β was conducted, and the shaded bars represent a case in which the induction of TGF- β was conducted. In the above experiment, cells after transfection were 45 cultured for 20 hours in the presence or absence of human TGF- β 1 (30 ng/ml) to prepare an extract from the cells, and luciferase was measured as described in H. Shibuya et al., Mol. Cell. Biol. Vol. 14, 5812 (1994). In the graph of Fig. 2, relative luciferase activities of cells transformed by a vector (containing no TAK1 gene) are shown with the luciferase activity in the absence of TGF- β 1 induction being set as 1. The results of bar graphs represent the mean of results of triplicate runs per experiment.
- 50 [0053] In order to confirm that the above results are mediated by the kinase activity of TAK1, a catalytically inactive TAK1 Δ N-K63W was constructed. This was carried out by site-directed mutagenesis using PCR. In this vector, lysine at position 63 in the ATP binding site has been replaced with tryptophan. The mutation is expected to inactivate the kinase activity and signaling activity of TAK1 Δ N. When TAK1 Δ N-K63W is co-transfected with p800neoLUC, the ability of constitutively stimulating the expression of the PAI-1 gene was lost (Fig. 2). These results suggest that the kinase activity 55 of TAK1 Δ N is required for the TGF- β -independent expression of PAI-1 gene. Furthermore, a kinase-negative TAK1 Δ N caused the partial reduction of TGF- β -induced expression. These results suggest that TAK1 may act as a mediator of a TGF- β -mediated route of signal transduction.
- [0054] In order to obtain direct evidence that TAK1 functions in the TGF- β -mediated route of signal transduction, it

was determined whether the treatment of cells with TGF- β could activate the kinase activity of TAK1. For the identification of an appropriate foreign substrate, an in vitro kinase reaction was conducted for TAK1 that was immunoprecipitated from yeast cells expressing TAK1 labeled with a hemagglutinin (HA) epitope (TAK1-HA) (a DNA sequence encoding an epitope recognized by anti-HA monoclonal antibody 12CA5 was ligated by PCR reaction to the 3'-terminal frame of DNA encoding TAK1).

[0055] The results of the immune complex kinase determination indicate that the activated form of TAK1 can phosphorylate and activate the XMEK2/SEK1 subfamily of MAPKK (B.M. Yasher et al., Nature Vol. 372, 794 (1994)). On the other hand, the phosphorylation of the original MAPKK-MEK1 (E. Nishida et al., Trends Biochem. Sci., 128 (1993); K.J. Blumer et al., ibid Vol. 19, 286 (1994); R.J. Davis, ibid Vol. 19, 470 (1990); C.L. Marchall, Cell, Vol. 80, 179 (1995)), histone and myelin basic protein was not detected. Thus, TAK1 kinase activity can be measured for its ability of activating XMEK2 in vitro.

[0056] Constructs for the expression of HA epitope-labeled TAK1 (HA-TAK1) were made in the following manner. A synthetic oligonucleotide encoding an HA epitope Tyr-Pro-Tyr-Asp-Val-Pro-Asp-Tyr-Ala (SEQ ID NO: 4) that is recognized by a monoclonal antibody 12CA5 was cloned into a Sall site (+3 position from the ATG codon) and an EcoRI site of pBS-TAK1 to construct pBS-HA-TAK1. pEF-MSS1 was cleaved with EcoRI and Sall, to which was inserted an EcoRI-Xhol fragment from pBS-HA-TAK1 to construct pEF-HA-TAK1.

[0057] In order to construct pBS-HA-TAK1 Δ N, pNV11-HU11 was digested with Xhol and HindIII. The fragment obtained was isolated and inserted into a HinclI-HindIII site of pBS-HA-TAK1. pEF-MSS1 was cleaved with EcoRI and Sall, into which was inserted a PstI-Xhol fragment from pBS-HA-TAK1 Δ H to construct pBS-HA-TAK1 Δ N. Both of these constructs have two copies of the N-terminal HA epitope expressed from the EF promoter.

[0058] These constructs pEF-HA-TAK1 or pEF-HA-TAK1 Δ H were transiently transfected to the MC3T3-E1 mouse osteoblast (S. Ohta et al., FEBS Lett. Vol. 314, 356 (1992)). After stimulation by TGF- β 1, the expressed HA-TAK1 was isolated by immunoprecipitation, and its activity was measured by the coupled kinase assay (S. Matsuda et al., J. Biol. Chem. Vol. 270, 12969 (1995)).

[0059] Thus, the transfected cells were treated with TGF- β 1 (20 ng/ml) or BMP-4 (100 ng/ml) for 0 minute (untreated) to 30 minutes. The cells were scraped into a buffer solution (S. Matsuda et al., J. Biol. Chem. Vol. 270, 12781 (1995); T. Moriguchi et al., J. Biol. Chem. Vol. 270, 12969 (1995)), and the cellular extract was centrifuged at 15,000 \times g for 10 minutes. The supernatant thus obtained was subjected to immunoprecipitation by anti-HA antibody. Thus, 300 μ l aliquots of the above supernatant were mixed with 20 μ l of antibody or 20 μ l of protein A Sepharose, and the immune complex was washed twice with PBS, which was then used for kinase measurement (S. Matsuda et al., J. Biol. Chem. Vol. 270, 12781 (1995); T. Moriguchi et al., J. Biol. Chem. Vol. 270, 12969 (1995)).

[0060] Activity was expressed as an incremental multiple relative to the HA-TAK1 activity of non-stimulated cells. The activity of immunoprecipitated TAK1 was measured by its ability of activating recombinant XMEK2/SEK1 (S. Matsuda et al., J. Biol. Chem. Vol. 270, 12781 (1995); T. Moriguchi et al., J. Biol. Chem. Vol. 270, 12781 (1995)).

[0061] It has already been confirmed that HA-TAK1 does not directly phosphorylate KN-p38/MPK2. In accordance with the immunoblotting of each immunoprecipitate by anti-HA antibody, almost identical amounts of HA-TAK1 were recovered at each point of immunoprecipitation.

[0062] The result of the above experiment indicated that TAK1 kinase activity started to rise within five minutes after stimulation by TGF- β , reached a peak at 10 minutes, and returned to an almost baseline level within 30 minutes (Fig. 3). Moreover, TGF- β 1 stimulated TAK1 kinase activity in a dose-dependent manner (Fig. 4). Then, it was determined whether TAK1 could be activated by BMP, a member of the TGF- β superfamily (A.H. Reddi et al., Curr. Opin. Genet. Dev. Vol. 4, 737 (1994)), or epithelial growth factor (EGF). Interestingly, BMP-4 also activated TAK1 kinase in a time- and dose-dependent manner (Fig. 4).

[0063] On the other hand, the activation of TAK1 was not observed in the cells treated with EGF. It is believed that the lack of TAK1 induction by EGF is not because MC3T3-E1 cells do not respond to EGF but because EGF signals are not mediated by TAK1. This is also apparent from the fact that EGF induces the expression of fos in MC3T3-E1 cells. Taken together, these data indicate that TAK1 is activated by the TGF- β superfamily.

[0064] TAK1 Δ N can activate the expression of PAI-1 gene independently of TGF- β (Fig. 2), which suggests that TAK1 Δ N protein has an enhanced kinase activity even in the absence of TGF- β treatment in the cell. In order to study this possibility, TAK1 Δ N labeled with the HA epitope (HA-TAK1 Δ N) (see above) was transiently transfected to MC3T3-E1 cells, and the activity of TAK1 Δ N was determined by the immunocomplex kinase measurement. Thus, MC3T3-E1 cells were transfected by pEF-HA-TAK1 Δ N, and from the transfected cells HA-TAK1 Δ N was immunoprecipitated as described above and its activity was measured.

[0065] All the data are expressed as an incremental multiple from the HA-TAK1 activity of non-stimulated cells.

[0066] As shown in Fig. 4, TAK1 Δ N protein has a considerably higher inherent kinase activity supporting the hypothesis that the TAK1 Δ N lacking the 22 amino acid residues at the N-terminal are constitutively active.

Example 5. Construction of cDNA library

[0067] From human T-cell cell line Jurkat cells, poly(A)RNA was prepared and cDNA was synthesized in a conventional method. This was inserted into a yeast expression vector pNV7 (Ninomiya-Tsuji, J. et al., Proc. Natl. Acad. Sci. U.S.A. 88, 9006-9010 (1991)) downstream of TDH3 promoter to construct a cDNA library.

Example 6. Screening of cDNA library

[0068] A mutant *Saccharomyces cereviceae* lacking the activity of Ssk2/Ssk22 and Sho1 that act in the signal transduction system of high osmotic pressure stress can grow in the YEPD medium (Yeast extract 10 g/l, tryptone 20 g/l, glucose 20 g/l), but not in a medium with 1M sorbitol added thereto (T. Maeda et al., Science, 269, 554 (1995)). Therefore, by introducing cDNA into this mutant followed by screening, a cDNA that can complement the deficient Ssk2/Ssk22 activity can be isolated.

[0069] In fact, a *Saccharomyces cereviceae* strain deficient in the Ssk2/Ssk22 and Sho1 activity described in the above reference (ssk2Δ, ssk22Δ, sho1Δ) was transformed with pNV11-HU11 (mouse TAK1ΔN) obtained in Example 2. The transformant was plated to the YEPD plate containing 1M sorbitol and was incubated at 30°C for 30 minutes. As a result, the yeast transformed with pNV11-HU11 grew even under a high osmotic pressure stress. This confirmed that the screening system is effective.

[0070] Then, this *Saccharomyces cereviceae* strain (ssk2Δ, ssk22Δ, sho1Δ) was transformed with the cDNA library constructed in Example 5, and was screened under a high osmotic pressure stress (incubated at 30°C in a YEPD medium containing 1M sorbitol). As a result, one positive clone pNV7-hTAK1 was obtained. cDNA contained in this clone was amplified using the PRISM Dye Terminator Cycle Sequencing kit (manufactured by Perkin Elmer) and the base sequence thereof was determined. The base sequence and the corresponding amino acid sequence were as set forth in SEQ ID NO: 5. The base sequence of this cDNA had a 92% homology with that of mouse TAK1, and the amino acid sequence encoded by the cDNA had a 99% homology with that of mouse TAK1. Comparisons of the base sequence of mouse TAK1 and human TAK1 are shown in Fig. 5 through Fig. 9, and those of the amino acid sequence are shown in Fig. 10 and Fig. 11.

[0071] Human TAK1 cDNA was subcloned into pUC19 that had been digested with Sall to obtain a plasmid pHAK1 containing the full-length cDNA of human TAK1. *E. coli* having the plasmid pHAK1 was internationally designated as Escherichia coli JM109 (pHAK1) under the provisions of the Budapest Treaty on July 19, 1996 with the National Institute of Bioscience and Human Technology, the Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as FERM BP-5598.

Deposit of microorganisms

[0072] The following microorganisms were deposited with the Patent Microorganism Depository in the National Institute of Bioscience and Human Technology, the Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, and were assigned the following accession numbers:

Organism: *Escherichia coli* MC1061/P3 (pEF-TAK1)
Date deposited: September 28, 1995
Accession number: FERM BP-5246
Organism: *Escherichia coli* MC1061/P3 (pEF-TAK1ΔN)
Date deposited: September 28, 1995
Accession number: FERM BP-5245
Organism: *Escherichia coli* JM109 (pHAK1)
Date deposited: July 19, 1996
Accession number: FERM BP-5598

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SEQUENCE LISTINGS

5 SEQ ID NO: 1
Sequence Length: 2443
10 Sequence Type: Nucleic acid
Strandedness: Single
15 Topology: Linear
Molecular Type: cDNA
Sequence
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CCACGGAGCG CCGGGCGGGCG GGCTGCGGGG CTCCGGGCTG AAGGGCGCTG CGCGAGCCGG 120
25 AGGGCGGGCG CGGCCCGCGG GGCGCCCGGG GGGATC ATG TCG ACA GCC TCC GCC 174
Met Ser Thr Ala Ser Ala
1 5
30 GCC TCG TCC TCC TCC TCG TCT TCT GCC AGT GAG ATG ATC GAA GCG CCG 222
Ala Ser Ser Ser Ser Ser Ser Ala Ser Glu Met Ile Glu Ala Pro
35 10 15 20
TCG CAG GTC CTG AAC TTC GAA GAG ATC GAC TAC AAG GAG ATC GAG GTG 270
Ser Gln Val Leu Asn Phe Glu Glu Ile Asp Tyr Lys Glu Ile Glu Val
40 25 30 35
GAA GAG GTT GTC GGA AGA GGA GCT TTT GGA GTA GTT TGC AAA GCT AAG 318
Glu Glu Val Val Gly Arg Gly Ala Phe Gly Val Val Cys Lys Ala Lys
45 40 45 50
TGG AGA GCA AAA GAT GTC GCT ATT AAA CAG ATA GAA AGT GAG TCT GAG 366
Trp Arg Ala Lys Asp Val Ala Ile Lys Gln Ile Glu Ser Glu Ser Glu
50 55 60 65 70

EP 0 919 621 A1

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	CCT AAC ATT GTC AAG TTG TAC GGA GCC TGC CTG AAT CCA GTA TGT CTT		462
10	Pro Asn Ile Val Lys Leu Tyr Gly Ala Cys Leu Asn Pro Val Cys Leu		
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15	GTG ATG GAA TAT GCA GAG GGG GGC TCA TTG TAT AAT GTG CTG CAT GGT		510
	Val Met Glu Tyr Ala Glu Gly Ser Leu Tyr Asn Val Leu His Gly		
	105	110	115
20	GCT GAA CCA TTG CCT TAC TAC ACT GCT GCT CAT GCC ATG AGC TGG TGT		558
	Ala Glu Pro Leu Pro Tyr Tyr Thr Ala Ala His Ala Met Ser Trp Cys		
	120	125	130
25	TTA CAG TGT TCC CAA GGA GTG GCT TAC CTG CAC AGC ATG CAG CCC AAA		606
	Leu Gln Cys Ser Gln Gly Val Ala Tyr Leu His Ser Met Gln Pro Lys		
	135	140	145
30	GCG CTG ATT CAC AGG GAC CTC AAG CCT CCA AAC TTG CTG CTG GTT GCA		654
	Ala Leu Ile His Arg Asp Leu Lys Pro Pro Asn Leu Leu Val Ala		
	155	160	165
35	GGA GGG ACA GTT CTA AAA ATC TGC GAT TTT GGT ACA GCT TGT GAC ATC		702
	Gly Gly Thr Val Leu Lys Ile Cys Asp Phe Gly Thr Ala Cys Asp Ile		
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	CAA ACA CAC ATG ACC AAT AAT AAA GGG AGT GCT GCT TGG ATG GCG CCT		750
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EP 0919 621 A1

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50 GAC ACT ATT AAA CGC TTG GAG TCA AAA CTG TTG AAA AAC CAG GCA AAG	1230
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EP 0 919 621 A1

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15	GAC ATG TCT GAA ATA GAA GCC AGG ATC GTG GCG ACT GCA GGT AAC GGG	1374
	Asp Met Ser Glu Ile Glu Ala Arg Ile Val Ala Thr Ala Gly Asn Gly	
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25	CCT GGT CAG GTG AGC AGC CGG TCA TCC AGC CCT AGT GTC AGA ATG ATC	1470
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30	ACT ACC TCA GGA CCA ACC TCA GAG AAG CCA GCT CGC AGT CAC CCA TGG	1518
	Thr Thr Ser Gly Pro Thr Ser Glu Lys Pro Ala Arg Ser His Pro Trp	
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	Thr Pro Asp Asp Ser Thr Asp Thr Asn Gly Ser Asp Asn Ser Ile Pro	
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	ATG GCG TAT CTT ACA CTG GAT CAC CAG CTA CAG CCT CTA GCG CCG TGC	1614
	Met Ala Tyr Leu Thr Leu Asp His Gln Leu Gln Pro Leu Ala Pro Cys	
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EP 0 919 621 A1

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SEQ ID NO: 2
50 Sequence Length: 16
Sequence Type: Nucleic acid

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Strandedness: Single
Topology: Linear
Molecular Type: Synthetic DNA
Sequence
AATTGCCAC CATGGC 16
SEQ ID NO: 3
Sequence Length: 16
Sequence Type: Nucleic acid
Strandedness: Single
Topology: Linear
Molecular Type: Synthetic DNA
Sequence
TCGAGCCATG GTGGCG 16
SEQ ID NO: 4
Sequence Length: 27
Sequence Type: Amino acid
Topology: Linear
Molecular Type: Peptide
Sequence
Tyr Pro Tyr Asp Val Pro Asp Tyr Ala
1 5
SEQ ID NO: 5
Sequence Length: 2656
Sequence Type: Nucleic acid
Strandedness: Single
Topology: Linear
Molecular Type: cDNA

EP 0 919 621 A1

Sequence

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GGGTCTCACC CGGATTGTCC GGGTGGCACC GTTCCCCGCC CCACCGGGCG CCGCGAGGGA 180
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40 CAG TTA TCC CGT GTG AAC CAT CCT AAT ATT GTA AAG CTT TAT GGA GCC 467
Gln Leu Ser Arg Val Asn His Pro Asn Ile Val Lys Leu Tyr Gly Ala
45 80 85 90 95
TGC TTG AAT CCA GTG TGT CTT GTG ATG GAA TAT GCT GAA GGG GGC TCT 515
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EP 0 919 621 A1

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	GCC CAC GCA ATG AGT TGG TGT TTA CAG TGT TCC CAA GGA GTG GCT TAT	611		
10	Ala His Ala Met Ser Trp Cys Leu Gln Cys Ser Gln Gly Val Ala Tyr			
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	CTT CAC AGC ATG CAA CCC AAA GCG CTA ATT CAC AGG GAC CTG AAA CCA	659		
15	Leu His Ser Met Gln Pro Lys Ala Leu Ile His Arg Asp Leu Lys Pro			
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20	CCA AAC TTA CTG CTG GTT GCA CGG GGG ACA GTT CTA AAA ATT TGT GAT	707		
	Pro Asn Leu Leu Leu Val Ala Gly Gly Thr Val Leu Lys Ile Cys Asp			
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25	TTT GGT ACA GCC TGT GAC ATT CAG ACA CAC ATG ACC AAT AAC AAG GGG	755		
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30	AGT GCT GCT TGG ATG GCA CCT GAA GTT TTT GAA GGT AGT AAT TAC AGT	803		
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35	GAA AAA TGT GAC GTC TTC AGC TGG GGT ATT ATT CTT TGG GAA GTG ATA	851		
	Glu Lys Cys Asp Val Phe Ser Trp Gly Ile Ile Leu Trp Glu Val Ile			
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40	ACG CGT CGG AAA CCC TTT GAT GAG ATT GGT GGC CCA GCT TTC CGA ATC	899		
	Thr Arg Arg Lys Pro Phe Asp Glu Ile Gly Gly Pro Ala Phe Arg Ile			
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	ATG TGG CCT GTT CAT AAT GGT ACT CGA CCA CCA CTG ATA AAA AAT TTA	947		
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EP 0 919 621 A1

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10	Ser Gln Arg Pro Ser Met Glu Glu Ile Val Lys Ile Met Thr His Leu			
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15	Met Arg Tyr Phe Pro Gly Ala Asp Glu Pro Leu Gln Tyr Pro Cys Gln			
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	TTG GGA GCC TCC CAT GGG AGC AGT GTG GAG AGC TTG CCC CCA ACC TCT	1331		
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EP 0919 621 A1

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15 Ser Pro Ser Val Arg Met Ile Thr Thr Ser Gly Pro Thr Ser Glu Lys
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CCA ACT CGA AGT CAT CCA TGG ACC CCT GAT GAT TCC ACA GAT ACC AAT 1571
20 Pro Thr Arg Ser His Pro Trp Thr Pro Asp Asp Ser Thr Asp Thr Asn
450 455 460
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30 CTA CAG CCT CTA GCA CCG TGC CCA AAC TCC AAA GAA TCT ATG GCA GTG 1667
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40 500 505 510
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SEQUENCE LISTING

EP 0 919 621 A1

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50
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EP 0 919 621 A1

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EP 0 919 621 A1

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EP 0 919 621 A1

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EP 0 919 621 A1

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EP 0 919 621 A1

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EP 0919 621 A1

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EP 0919 621 A1

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EP 0919 621 A1

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	Leu Gly Ala Ser His Gly Ser Ser Val Glu Ser Leu Pro Pro Thr Ser		
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			415
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45	Ser Pro Ser Val Arg Met Ile Thr Thr Ser Gly Pro Thr Ser Glu Lys		
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	cca act cga agt cat cca tgg acc cct gat gat tcc aca gat acc aat		1571
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EP 0919 621 A1

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EP 0 919 621 A1

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Val Val Cys Lys Ala Lys Trp Arg Ala Lys Asp Val Ala Ile Lys Gln

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Ile Glu Ser Glu Ser Glu Arg Lys Ala Phe Ile Val Glu Leu Arg Gln

65 70 75 80

50 Leu Ser Arg Val Asn His Pro Asn Ile Val Lys Leu Tyr Gly Ala Cys

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EP 0 919 621 A1

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Tyr Asn Val Leu His Gly Ala Glu Pro Leu Pro Tyr Tyr Thr Ala Ala

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15 His Ser Met Gln Pro Lys Ala Leu Ile His Arg Asp Leu Lys Pro Pro

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Gly Thr Ala Cys Asp Ile Gln Thr His Met Thr Asn Asn Lys Gly Ser

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Ala Ala Trp Met Ala Pro Glu Val Phe Glu Gly Ser Asn Tyr Ser Glu

195 200 205

30 Lys Cys Asp Val Phe Ser Trp Gly Ile Ile Leu Trp Glu Val Ile Thr

210 215 220

35 Arg Arg Lys Pro Phe Asp Glu Ile Gly Gly Pro Ala Phe Arg Ile Met

225 230 235 240

Trp Ala Val His Asn Gly Thr Arg Pro Pro Leu Ile Lys Asn Leu Pro

40 245 250 255

Lys Pro Ile Glu Ser Leu Met Thr Arg Cys Trp Ser Lys Asp Pro Ser

260 265 270

45 Gln Arg Pro Ser Met Glu Glu Ile Val Lys Ile Met Thr His Leu Met

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EP 0919 621 A1

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EP 0 919 621 A1

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EP 0 919 621 A1

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EP 0919 621 A1

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15 Cys Lys Lys Gln Leu Glu Val Ile Arg Ser Gln Gln Gln Lys Arg Gln
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20 Gly Thr Ser

25

Claims

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1. A polypeptide having a kinase activity that is activated by transforming growth factor (TGF)- β , said polypeptide comprising an amino acid sequence from Ser at position 23 to Ser at position 579 set forth in SEQ ID NO: 5.
2. A polypeptide having a kinase activity that is activated by TGF- β , said polypeptide comprising an amino acid sequence from Met at position 1 to Ser at position 579 set forth in SEQ ID NO: 5.
3. DNA encoding a polypeptide having a kinase activity that is activated by TGF- β , said polypeptide comprising an amino acid sequence from Ser at position 23 to Ser at position 579 set forth in SEQ ID NO: 5.
- 35 4. DNA according to claim 3 having a nucleotide sequence from T at position 249 to A at position 1919 set forth in SEQ ID NO: 5.
5. DNA encoding a polypeptide having a kinase activity that is activated by TGF- β , said polypeptide comprising an amino acid sequence from Met at position 1 to Ser at position 579 set forth in SEQ ID NO: 5.
- 40 6. DNA according to claim 5 having a nucleotide sequence from A at position 183 to A at position 1919 set forth in SEQ ID NO: 5.
7. A vector comprising DNA according to any of claims 3 to 6.
- 45 8. A host cell transformed with a vector comprising DNA according to any of claims 3 to 6.
9. A method for producing a polypeptide having a kinase activity that is activated by TGF- β , which method comprises culturing a host cell transformed with a vector comprising DNA according to any of claims 3 to 6 and then recovering the product from the culture.
- 50 10. A polypeptide having a kinase activity that is activated by TGF- β , said polypeptide being produced by the method according to claim 9.
11. A kinase that is activated by TGF- β , said kinase comprising an amino acid sequence from Ser at position 23 to Ser at position 579 set forth in SEQ ID NO: 5.
- 55 12. A fusion protein of a protein according to any of claims 1, 2, 10, and 11, and another protein.

Fig. 1

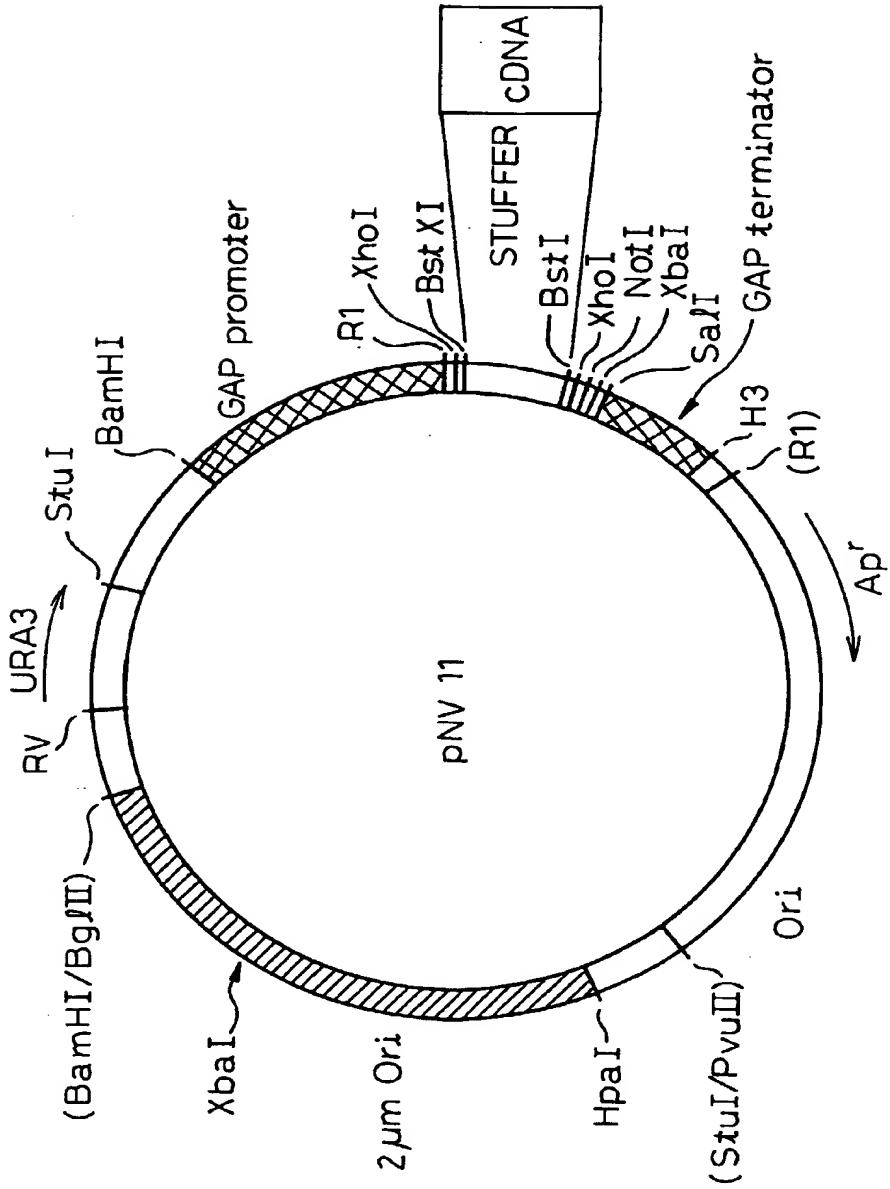


Fig.2

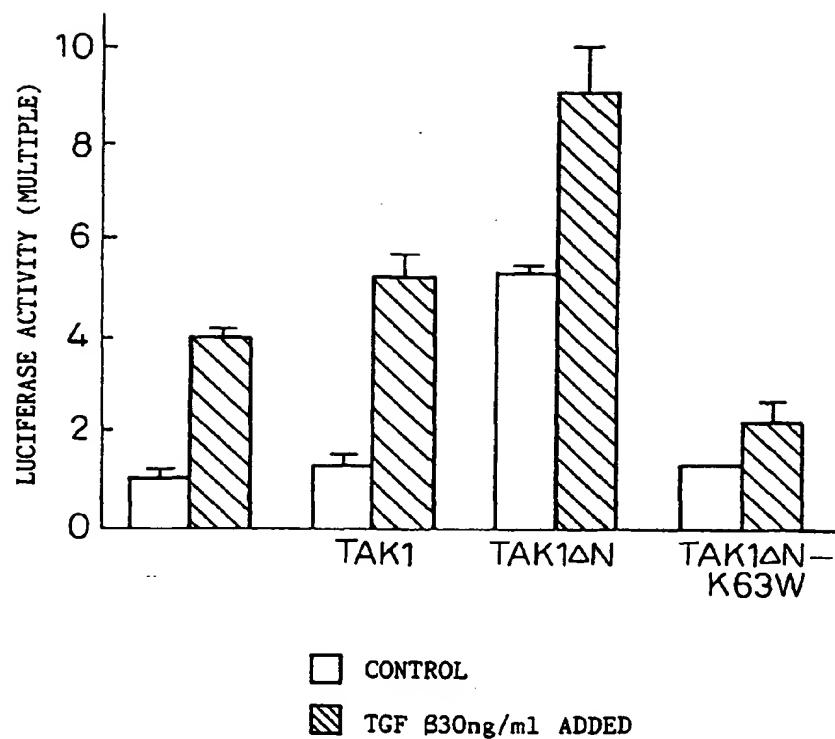


Fig.3

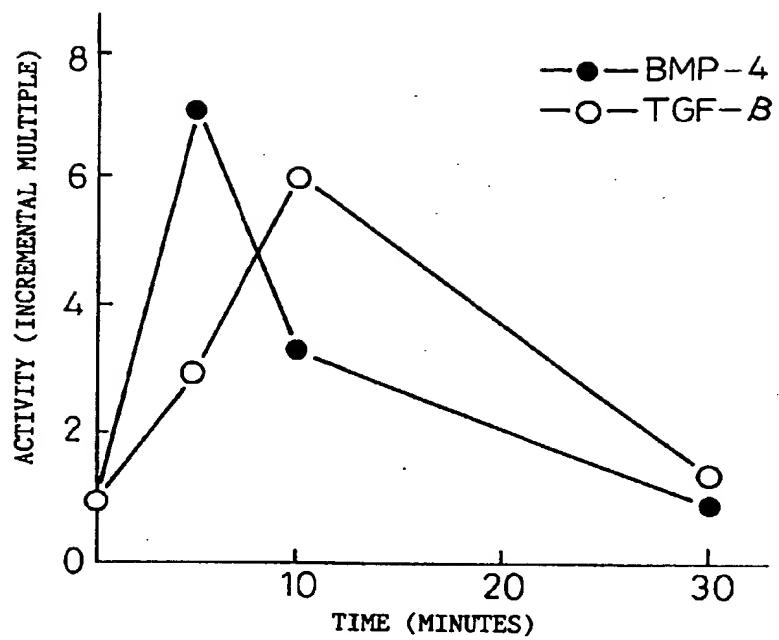


Fig.4

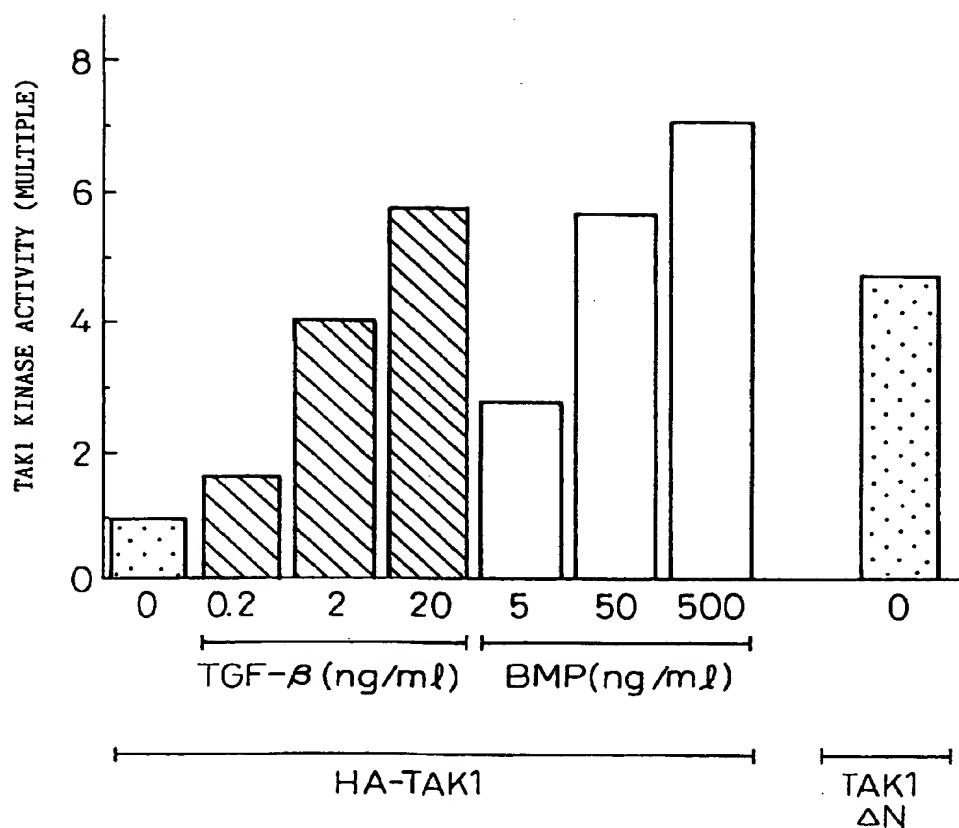


Fig. 5

EP 0 919 621 A1

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1' ATGTCTACGCCCTCGCCGCTCTCCCTCGCTTCGGCCGGTGAATGATCGAA
** * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
1" ATGTCGACAGCCCTCCGGCCTCGCTCCCTCGCTTCGGCCAGTGATGATCGAA

61' GCCCCTCCAGGTCCAACTTGAAGAGATCGACTCAAGGAGATCGAGGTGGAAGAG
** * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
61" GCGCCGTCGAGGTCCCTGAACCTCGAAGAGATCGACTCAAGGAGATCGAGGTGGAAGAG

121' GTGTGGAAAGAGGCCCTGGAGTTGAAAGCTCAAGGAGATCGACTCAAGGAGATCGAGGTGATGTT
** * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
121" GTGTGGAAAGAGGCCCTGGAGTAGTTGCAAAGCTAAAGTGGAGAGCAAAAGATGTC

181' GCTATTAAACAAATAGAAAAGTGAATCTGAGAGGAAAGCGTTATTGTAGAGCTTCGGCA
** * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
181" GCTATTAAACAGATAGAAAAGTGAAGCTCTGAGAGGAAGGCCCTCATTTGGAGCTCCGGCA

241' TTATCCCGTGTGAACCATCCTAAATTGTAAAGCTTTATGGAGCCCTGCTTGATCCAGTG
** * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
241" TTGTCCGGTGTGAACCATCCTAACATTGCAAGTGTACGGAGCCCTGATCCAGTA

301' TGTCTTGTGATGGAATATGCTGAAGGGGGCTTTATAATGTGCTGCATGGCTGAA
** * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
301" TGTCTTGTGATGGAATATGCAAGGGGGCTCATTTGATAATGTGCTGCATGGCTGAA

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卷之六

EP 0 919 621 A1

Fig. 7

EP 0 919 621 A1

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661' GAAGTGTACGGAAACCTTGTAGAGATTGGTGGCCAGCTTCCGAATCATG
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * *
661" GAAGTGTACACGCCAACCTCGTGTAGATCGTGGCCAGCTTCAAGATCATG
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
721' TGGGCTGTTCATATGGTACTCGACCACTGATAAAAATTACCTAACGCCATTGAG
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
721" TGGGCTGTTCATATGGCACTCGACCACTGATAAAAATTACCTAACGCCATTGAG
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
781' AGCCTGATGACTCGTTGGTCTAAAGATCCTCCAGGCCCTCAATGGAGGAATT
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
781" AGCTTGATGACACGCTGTGGTCTAAGGACCCATCTAGGCCCTCAATGGAGGAATT
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
841' GTGAAAATTAATGACTCACTTGATGGGTACTTCCAGGAGCAGATGAGCCATTACAGTAT
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
841" GTGAAAATTAATGACTCACTTGATGGGTACTTCCAGGAGGGATGAGCCATTACAGTAT
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
901' CCTTGTCAGTATTCAAGATGAAGGACAGCAACTCTGCCACCACTGACAGGCTCATG
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
901" CCTTGTCAGTACTCTGATGAAGGCAGAGCAACTCACGCCACAGCACAGGCTCGTTCATG
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
961' GACATTGCTCTACAATAACGAGTAACAAAAGTGACACTAATATGGAGCAAGTCCCTGCC
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
961" GACATTGCTCTACAATAACGAGTAATAAAGTGACACAATAATGGAAACAAATATGGAGCAAGTCCCTGCC
***** * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

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卷之二

EP 0 919 621 A1

1021 '	ACAAATGATACTATTAAAGCGCTTAAAGCTGGGAGCAGTTGGAGCTCCCATGGGAGCTGGAGCTGGGGCTGCC
1021 "	ACAAACGACACTATTAAACCGCTTGGAGTCAAACTGTGAAAAACCCAGGCAAAAGCAACAG
1081 '	AGTGAATCTGGACGTTTAAGCTTGGGAGCAGTTGGAGCTCCCATGGGAGCTGGAGCTGGGGCTGCC
1081 "	AGTGAATCTGGACGCCTGAGCTGGGAGCCCTCTCGTGGAGGCAATGTGCTGAGAGCTGGGGCTGCC
1141 '	CCAACCTCTGAGGGCAAGAGGATGAGTGTGACATGTCTGAAATAGAACGCTAGGATCTGGCC
1141 "	CCCACCTCCGAGGGCAAGAGGATGAGTGTGACATGTCTGAAATAGAACGCTAGGATCTGGCC
1201 '	GCAACCACAGGCAACGGCACAGGCCAAGACGTTAGATCCATCCAAGACTGACTGTAACTGG
1201 "	GCGACTGCAAGGTAACGGGAACCAAGGGTAGATCCATCCAAGACTGACTGTACTGGG
1261 '	ACAGAACCTGGTCAGGTGAGGCACTGTCAGTCCACTGTCAGAATGATTAATCACC
1261 "	ACAGAACCTGGTCAGGTGAGCAGCCGTATCCAGCCCTAGTGTCAAGAATGATCACTACC
1321 '	TCAGGACCAACCTCAGAAAAGCCAACTCGAACGCTGACATCCATGGACCCCTGTGATGATTCCACAA
1321 "	TCAGGACCAACCTCAGAAAAGCCAGCTGGCACTGGACCCATGGACCCCTGTGATGATTCCACAA

卷之三

EP 0 919 621 A1

1381' GATACCAATGGATCAGATAACTCCATCCCATTGGCTTACACTGGATCACCAACTA
1381" GACACCAATGGCTCAGATAACTCCATCCCATTGGGTATCTTACACTGGATCACCACTA

1441' CAGCCTCTAGGCCAACCGTGCCTAAACTCCAAGAATCTATGGCAGTGTGTTAACAGGCTATTGT
1441" CAGCCTCTAGGCCGTGCCTAAACTCCAAGAATCCATGGCAGTGTGTTAACAGGCTATTGT

1501' AAAATGGCACAAAGAAATAATATGAAAGTTCAACAGAAATTGGCATTTGTTTACAGAGAAAG
1501" AAAATGGCACAGGGATATATGAAAGTTCAACCGAAATGGCATTGTTACTACAGAGAAAG

1561' CAAGGAACACTAGTGCAGAACGGACAGGATGAAAAGGACAGCAAAATAACATCTGCCCTG
1561" CAAGGAACACTAGTGCAGAACGGACAGGATGAAAAGGACAGCAAAATAACATCTGCCCTG

1621' GTACAGGAACATAAAAAGCTTTAGATGAAAAACAAAAGCCTTCTACTTACTACAGCAA
1621" GTACAGGAACATAAAAAGCTTTAGATGAAAAACAAAAGCCTTCTACTTATTACAGCAA

1681' TGCACAAAAACAACTAGAGGTCTACAGAACGAGCAAGCAAGGCACTTCATGA
1681" TGCACAAAAACAACTAGAGGTCTACAGAACGAGCAAGGCACTTCATGA

Fig. 10

EP 0 919 621 A1

1' MSTASAASSSSSSAGEMIEAPSQVLNFEEDIKYKEIEVEEVVGRGAFGVVCKAKWRAKDV

1" MSTASAASSSSSSASEMIEAPSQVLNFEEDIKYKEIEVEEVVGRGAFGVVCKAKWRAKDV

61' AIKQIESESERKAFIGVELRQLSRVNHPNIVKLYGACLNPVCLVMEYAEGGSLYNVLHGAE

61" AIKQIESESERKAFIGVELRQLSRVNHPNIVKLYGACLNPVCLVMEYAEGGSLYNVLHGAE

121' PLPYYTAAHAMSWCLQCSQGVAYLHSMQPKALIHRDLKPPNLLVAGGTVLKICDFGTAC

121" PLPYYTAAHAMSWCLQCSQGVAYLHSMQPKALIHRDLKPPNLLVAGGTVLKICDFGTAC

181' DIQTHMTNNKGSAAMMAPEVFGSNYSEKCDVFSWGIILWEITRKPFDEIGGPAPRIM

181" DIQTHMTNNKGSAAMMAPEVFGSNYSEKCDVFSWGIILWEITRKPFDEIGGPAPRIM

241' WAVHNGTRPPLIKNLPKPIESLMTRCWSKDPQRPSMEIVKIMTHLMRYFPGADEPLQY

241" WAVHNGTRPPLIKNLPKPIESLMTRCWSKDPQRPSMEIVKIMTHLMRYFPGADEPLQY

Fig. 11

301' PCQYSDEGGQNSATSTGSFMDIASTNTSNKSDTNMEQVPATNDTIKRLESKLLKNQAKQQ

301" PCQYSDEGGQNSATSTGSFMDIASTNTSNKSDTNMEQVPATNDTIKRLESKLLKNQAKQQ

361' SESGRRLSLGASHGSSVESLPPTEGKRMSSADMSEIEARIAATTGNGPRRRSIQDLTVTG

361" SESGRRLSLGASRGSSSVESLPPTEGKRMSSADMSEIEARIVATAGNGPRRRSIQDLTVTG

421' TEPGQVSSSSSPSVRMITTSGPTSEKPTTRSHPWTPDDSTDNTGSDNSIPMAYLTLDHQI

421" TEPGQVSSSSSPSVRMITTSGPTSEKPARSHPWTPDDSTDNTGSDNSIPMAYLTLDHQI

481' QPLAPCPNSKESMAFEQHCKMMAQEYMKVQTEIALLQRKQELVAYELDQDEKDQQNTSRL

481" QPLAPCPNSKESMAFEQHCKMMAQEYMKVQTEIALLQRKQELVAYELDQDEKDQQNTSRL

541' VQEHKKKLDENKSLSTYYQQCKKKLEVIRSSQQKQRQGTS

541" VQEHKKKLDENKSLSTYYQQCKKKLEVIRSSQQKQRQGTS

EP 0 919 621 A1

INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP97/01050

A. CLASSIFICATION OF SUBJECT MATTER
Int. Cl⁶ C12N15/54, C12P21/02, C12N9/12, C12N1/21

According to international Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

Int. Cl⁶ C12N15/54, C12P21/02, C12N9/12, C12N1/21

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

CAS ONLINE, BIOSIS, WPI/WPI, L

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Science 270 1995 K. Yamaguchi et al. "Identification of a member of the MAPKKK Family as a potential mediator of TGF-β signal transduction" p. 2008-2011	1 - 12
Y	Hyuga Saito and others "New Molecular Genetics for Bio-Science (in Japanese)" (Nankodo) 1987 p. 235-236	12
A	Cell 80 1995 C.J. Marshall "Specificity of receptor tyrosine kinase signaling: transient versus sustained extracellular signal-regulated kinase activation" p. 179-185	1 - 12
A	Science 265 1994 J.F. Smothers et al. "Stimulatory effects of yeast and mammalian 14-3-3 proteins on the raf protein kinase" p. 1716-1719	1 - 12
A	Science 241 1988 Steven K. Hanks et al. "The protein kinase family: conserved features and	1 - 12

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Date of the actual completion of the international search
June 24, 1997 (24. 06. 97)Date of mailing of the international search report
July 8, 1997 (08. 07. 97)Name and mailing address of the ISA/
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INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP97/01050

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>deduced phylogeny of the catalytic domains" p. 42-52</p> <p>Nature 324 1986 Randall K. Saiki et al. "Analysis of enzymatically amplified β-globin and HLA-DQα DNA with allele-specific oligonucleotide probes" p. 163-166</p>	1 - 12

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European Patent
Office

SUPPLEMENTARY
EUROPEAN SEARCH REPORT

0919621

Application Number
EP 97 90 8525

DOCUMENTS CONSIDERED TO BE RELEVANT			CLASSIFICATION OF THE APPLICATION (Int.CI.6)									
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim										
T	<p>SAKURAI, H. ET AL.: "TGF-beta-Activated Kinase 1 Stimulates NF-kappaB Activation by an NF-kappaB-Inducing Kinase-Independent Mechanism" BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 243, no. 2, 13 February 1998 (1998-02-13), pages 545-549, XP000867678 * page 545, column 2, line 32 - page 546, column 1, line 14 * * page 546, column 1, line 60 - column 2, line 11 * * page 547; figure 1A *</p> <p>-----</p>	1-12	C12N15/54 C12P21/02 C12N9/12 C12N1/21									
			TECHNICAL FIELDS SEARCHED (Int.Cl.6)									
			C12N C12P									
1	<p>The supplementary search report has been based on the last set of claims valid and available at the start of the search.</p> <table border="1"> <tr> <td>Place of search BERLIN</td> <td>Date of completion of the search 22 February 2000</td> <td>Examiner Fuchs, U</td> </tr> <tr> <td colspan="3"> CATEGORY OF CITED DOCUMENTS <ul style="list-style-type: none"> <input checked="" type="checkbox"/> X : particularly relevant if taken alone <input checked="" type="checkbox"/> Y : particularly relevant if combined with another document of the same category <input checked="" type="checkbox"/> A : technological background <input checked="" type="checkbox"/> O : non-written disclosure <input checked="" type="checkbox"/> P : intermediate document </td> </tr> <tr> <td colspan="3"> <ul style="list-style-type: none"> <input checked="" type="checkbox"/> T : theory or principle underlying the invention <input checked="" type="checkbox"/> E : earlier patent document, but published on, or after the filing date <input checked="" type="checkbox"/> D : document cited in the application <input checked="" type="checkbox"/> L : document cited for other reasons <input checked="" type="checkbox"/> & : member of the same patent family, corresponding document </td> </tr> </table>			Place of search BERLIN	Date of completion of the search 22 February 2000	Examiner Fuchs, U	CATEGORY OF CITED DOCUMENTS <ul style="list-style-type: none"> <input checked="" type="checkbox"/> X : particularly relevant if taken alone <input checked="" type="checkbox"/> Y : particularly relevant if combined with another document of the same category <input checked="" type="checkbox"/> A : technological background <input checked="" type="checkbox"/> O : non-written disclosure <input checked="" type="checkbox"/> P : intermediate document 			<ul style="list-style-type: none"> <input checked="" type="checkbox"/> T : theory or principle underlying the invention <input checked="" type="checkbox"/> E : earlier patent document, but published on, or after the filing date <input checked="" type="checkbox"/> D : document cited in the application <input checked="" type="checkbox"/> L : document cited for other reasons <input checked="" type="checkbox"/> & : member of the same patent family, corresponding document 		
Place of search BERLIN	Date of completion of the search 22 February 2000	Examiner Fuchs, U										
CATEGORY OF CITED DOCUMENTS <ul style="list-style-type: none"> <input checked="" type="checkbox"/> X : particularly relevant if taken alone <input checked="" type="checkbox"/> Y : particularly relevant if combined with another document of the same category <input checked="" type="checkbox"/> A : technological background <input checked="" type="checkbox"/> O : non-written disclosure <input checked="" type="checkbox"/> P : intermediate document 												
<ul style="list-style-type: none"> <input checked="" type="checkbox"/> T : theory or principle underlying the invention <input checked="" type="checkbox"/> E : earlier patent document, but published on, or after the filing date <input checked="" type="checkbox"/> D : document cited in the application <input checked="" type="checkbox"/> L : document cited for other reasons <input checked="" type="checkbox"/> & : member of the same patent family, corresponding document 												

CLAIMS

1. A polypeptide having a kinase activity that is activated by transforming growth factor (TGF)- β , said polypeptide comprising an amino acid sequence from Ser at position 23 to Ser at position 579 set forth in SEQ ID NO: 5.
2. A polypeptide having a kinase activity that is activated by TGF- β , said polypeptide comprising an amino acid sequence from Met at position 1 to Ser at position 579 set forth in SEQ ID NO: 5.
3. DNA encoding a polypeptide having a kinase activity that is activated by TGF- β , said polypeptide comprising an amino acid sequence from Ser at position 23 to Ser at position 579 set forth in SEQ ID NO: 5.
4. DNA according to claim 3 having a nucleotide sequence from T at position 249 to A at position 1919 set forth in SEQ ID NO: 5.
5. DNA encoding a polypeptide having a kinase activity that is activated by TGF- β , said polypeptide comprising an amino acid sequence from Met at position 1 to Ser at position 579 set forth in SEQ ID NO: 5.
6. DNA according to claim 5 having a nucleotide sequence from A at position 183 to A at position 1919 set forth in SEQ ID NO: 5.
7. A vector comprising DNA according to any of claims 3 to 6.
8. A host cell transformed with a vector comprising DNA according to any of claims 3 to 6.
9. A method for producing a polypeptide having a kinase activity that is activated by TGF- β , which method comprises culturing a host cell transformed with a vector comprising DNA according to any of claims 3 to 6 and then recovering the product from the culture.
10. A polypeptide having a kinase activity that is activated by TGF- β , said polypeptide being produced by the method according to claim 9.
11. A kinase that is activated by TGF- β , said

- 32 -

kinase comprising an amino acid sequence from Ser at position 53 to Ser at position 179 set forth in SEQ ID NO: 5.

12. A fusion protein of a protein according to any
5 of claims 1, 2, 10, and 11, and another protein.

- [33] -

ABSTRACT

A TGF- β -activated kinase comprising an amino acid sequence from Met at position 1 to Ser at position 579 in
5 the amino acid sequence as set forth set forth in SEQ ID NO: 5, and DNA encoding the kinase.